

Chronic Hepatitis A With Persistent Viral Replication

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Hepatitis A virus (HAV) usually causes an acute self-limited illness. This report describes a patient with hepatitis A whose serum aminotransferase activities remained above normal and whose serum was persistently positive for immunoglobulin (Ig) M class anti-hepatitis A 31 months after the onset of hepatitis. Liver biopsy carried out 11 months after the onset of hepatitis showed histological changes consistent with chronic hepatitis of moderate severity. HAV RNA was detected by polymerase chain reaction (PCR) in feces collected at the time of the liver biopsy. Furthermore, the patient developed esophageal varices 25 months after the onset of hepatitis. We believe this to be the first reported case in which persistent replication of HAV is implicated in chronic hepatitis with the potential to develop into liver cirrhosis. © 1996 Wiley-Liss, Inc.

KEY WORDS: HAV RNA, chronic hepatitis, IgM-HA antibody

INTRODUCTION

It is generally believed that hepatitis A virus (HAV) causes acute self-limiting hepatitis that never leads to chronic hepatitis or a chronic carrier state. However, McDonald et al. [1989] reported the case of a young male with acute hepatitis A who was serologically positive for IgM anti-HAV for more than 1,000 days. Liver biopsy undertaken 2 years after the onset of hepatitis showed that he had chronic active hepatitis with progressive fibrosis. In that report, however, persistent HAV replication was not demonstrated in the patient and that chronic hepatitis caused by HAV is not certain.

In the present report we describe a case histologically confirmed to be chronic hepatitis with moderate severity and the potential to develop liver cirrhosis. Virological evidence is presented of persistent HAV replication for more than 6 months after the diagnosis of hepatitis, and no evidence of infection with other hepatitis viruses.

Method

Commercial kits (Abbott Laboratories, North Chicago, IL) were used to test for immunoglobulin (Ig) M antibody

to HAV (anti-HA), hepatitis B surface antigen (HBsAg), antibody to HBsAg (anti-HBs), antibody to HB core antigen (anti-HBc), HBeAg, antibody to HBeAg (anti-HBe), and antibody to hepatitis D virus (anti-HD). The presence of antibody to hepatitis C virus (anti-HCV) was determined by a second-generation commercial kit (Ortho Diagnostics, Tokyo, Japan). The presence of antibody to human immunodeficiency virus (anti-HIV) was tested by a commercial kit (Fujirebio, Tokyo, Japan). HCV RNA was tested by nested polymerase chain reaction (PCR), using two sets of primers deduced from the 5'-noncoding region of HCV [Okamoto et al., 1990]. Hepatitis B virus (HBV) DNA was tested by PCR using a set of primers deduced from the HBsAg region of HBV DNA.

HAV RNA was detected by the reverse transcription (RT)-PCR method previously reported [Yotsuyanagi et al., 1993]. Briefly, HAV RNA was amplified with two different sets of primers. One set of primers described by Cohen et al. [1987] was deduced from the 5'-noncoding region of the HAV genome. The other set of primers described by Jansen et al. [1990] was deduced from the VP3 region. PCR products were analyzed by electrophoresis in 6% polyacrylamide gels and by ethidium bromide staining.

Case

A 48-year-old male patient was admitted to Showa University Fujigaoka Hospital with marked jaundice and general malaise. There was no family history of liver disease, prior history of jaundice, nor known blood transfusion or intravenous drug abuse. The patient reported having eaten raw fish several times a month before admission. He complained of becoming fatigued easily, and had had high fever 7 days before admission. He consulted a family physician and was found to be jaundiced. On March 15, 1993 he was referred to our hospital and admitted.

Physical examination revealed marked jaundice and a mildly enlarged liver. On neurological examination the patient was alert and fully oriented. Hematological tests

Accepted for publication July 18, 1996.

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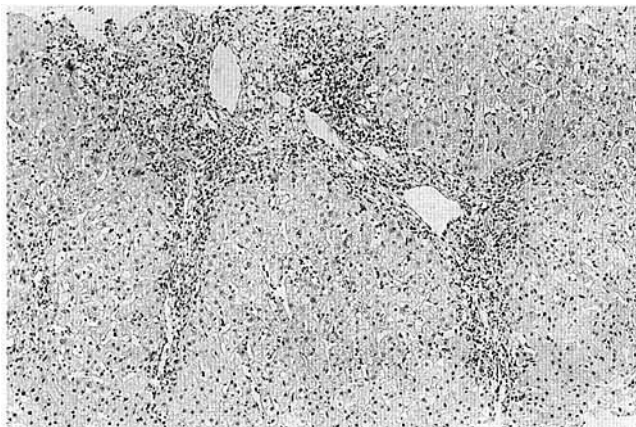


Fig. 2. Liver biopsy specimen obtained on February 24, 1994. Portal expansion and moderate interface hepatitis as well as slight lobular activity, consistent with chronic hepatitis of moderate severity. HE $\times 200$.

improving severe liver dysfunction. The treatment was of a limited value, however, in eradicating HAV.

We have treated six cases with fulminant hepatitis A who showed a prolonged elevation of IgM anti-HA and aminotransferase activities with IFN. The total amount of administered IFN in these six cases was 138 MU (105–267 MU) which was greater than that in the present case. IFN seemed effective in improving liver function and eradicating HAV in these patients because all survived, resulting in the final elimination of HAV. Although IFN seemed effective in improving liver function, the total amount may have been too small for the eradication of HAV in the present case.

Chronicity of hepatitis was established by liver biopsy performed 11 months after the onset of hepatitis. Aminotransferase activities fluctuated persistently with elevated IgM anti-HA level as late as 31 months after the onset of hepatitis. Furthermore, esophageal varices were observed 25 months after the onset of hepatitis. The presence of liver cirrhosis is not ascertained at present. Above observations imply, however, that hepatitis in this patient is not merely prolonged but has the potential to develop into liver cirrhosis.

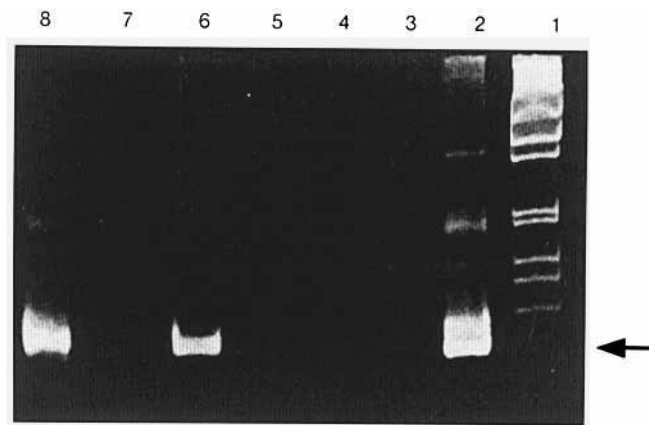


Fig. 3. Detection of HAV RNA by RT-PCR. **Lane 1:** Molecular weight markers. **Lanes 2–5:** RNA from patient's serum. **Lane 6:** Positive control obtained from HAV cultures. **Lane 7:** Negative control. **Lane 8:** RNA from a patient's feces. PCR products of HAV RNA have 272 base pairs in size.

REFERENCES

- Cohen JI, Ticehurst JR, Purcell RH, Bulukler-White A, Baroudy BM (1987): Complete nucleotide sequence of wild-type hepatitis A virus: Comparison with different strains of hepatitis A virus and other picornaviruses. *Journal of Virology* 61:50–59.
- Jansen RW, Siegel G, Lemon SM (1990): Molecular epidemiology of human hepatitis A virus defined by an antigen-capture polymerase chain reaction method. *Proceedings of the National Academy of Sciences USA* 87:2867–2871.
- Ludwig J (1993): The nomenclature of chronic active hepatitis: An obituary. *Gastroenterology* 105:274–278.
- McDonald GSA, Courtney MG, Shattock AG, Weir DG (1989): Prolonged IgM antibodies and histopathological evidence of chronicity in hepatitis A. *Liver* 9:223–228.
- Okamoto H, Okuda S, Sugiyama Y, Tanaka T, Sugai Y, Akahane Y, Machida A, Mishiri S, Yoshizawa H, Miyakawa Y, Mayumi M (1990): Detection of hepatitis C virus RNA by a two-stage polymerase chain reaction with two pairs of primers deduced from 5'-noncoding region. *Japanese Journal of Experimental Medicine* 60:215–222.
- Vallbracht A, Flehming B (1985): Elimination of persistent hepatitis A infection in cell culture by interferon. In Kircher H, Shellenkens H (eds): "The Biology of the Interferon System." New York: Elsevier, 1985, pp 339–345.
- Yoshida M, Inoue K, Sekiyama K (1994): Interferon for hepatitis A. *Lancet* 343:288–289.
- Yotsuyanagi H, Iino S, Koike K, Yasuda K, Hino K, Kurikawa K (1993): Duration of viremia in human hepatitis A viral infection as determined by polymerase chain reaction. *Journal of Medical Virology* 40:35–38.